

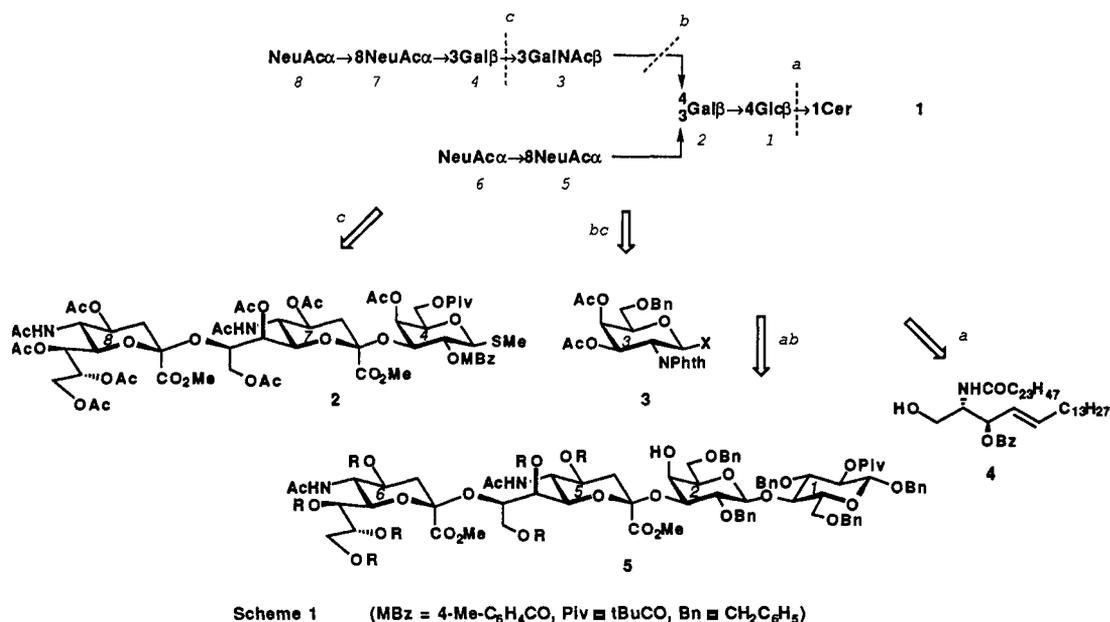
Experiments directed towards synthesis of complex glycosphingolipids: Ganglio-ganglioside GQ1b¹

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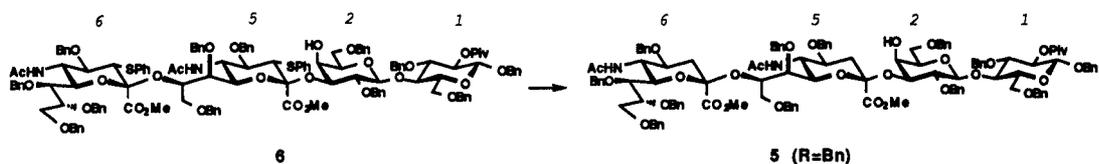
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Abstract: Synthesis of a properly protected octasaccharide **18** which could be regarded as a reasonable precursor for the synthesis of GQ1b **1** was carried out in a stereo- and regio-controlled manner.

Tetra-sialo ganglio-ganglioside GQ1b **1** was first isolated in 1967 from human brain² and then in 1972 from fish brain³. The structure of **1** was characterized around 1980 through chemical and enzymic degradation⁴ as well as by Mass spectral data⁵. The trophic effect of GQ1b on the nervous system has been shown by employing human neuroblastoma cell line⁶. Since structurally most simple ganglioside GM₃ was synthesized for the first time in 1985⁷, more efficient strategies for the synthesis of ganglio-gangliosides have been developed⁸. A total synthesis of GQ1b, however, still remains to be accomplished. In this paper, we highlight successful synthesis of a properly protected octasaccharide derivative **18** which may be regarded as a key intermediate for further conversion into GQ1b.

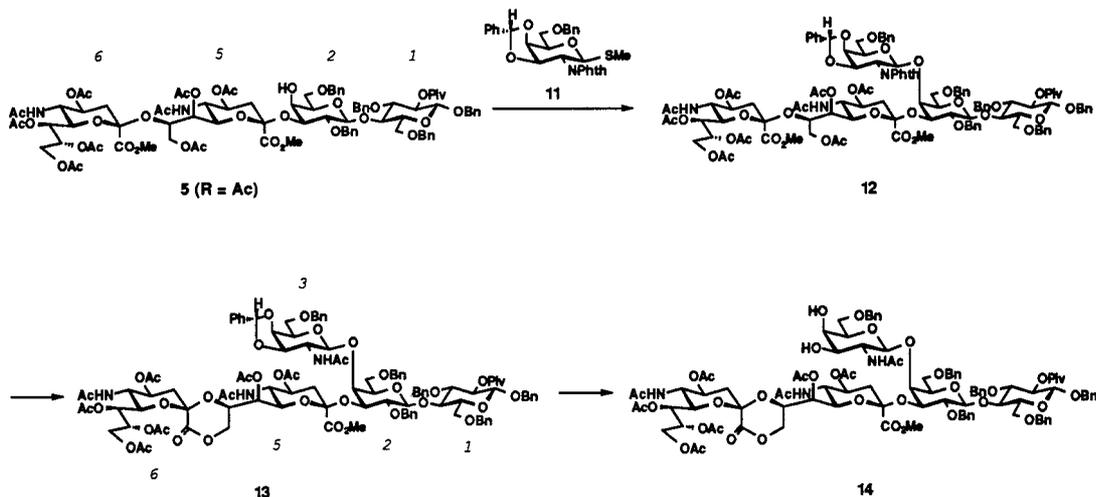


Aiming at a convergent-type synthesis of GQ1b **1**, strategic bond disconnection was planned as shown in scheme 1 which led to design completely protected building blocks **2**, **3**, **4**, and **5**. It is to be noted that 4-methylbenzoyl and pivaloyl groups at O-2 of monosaccharide residue **4** and **1** in compounds **2** and **5**, respectively, are required to serve as a stereocontrolling auxiliary⁹.



Scheme 2

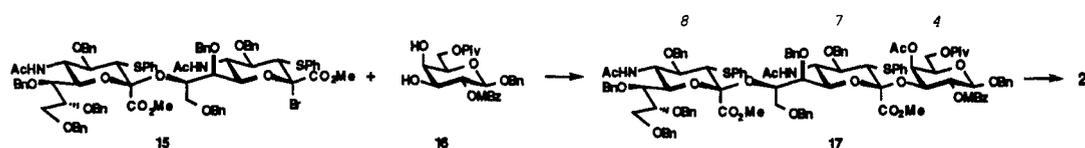
In order to study the reactivity of O-4 at residue **2** of the building block **5**, first we prepared **5** [R=Bn, R_F 0.37 in 4:1 PhMe-Me₂CO; δ_H 5.107 (dd, 8.0 and 9.5 Hz, H-2¹), 3.748 and 3.510 (2s, 2 x OMe), 2.816 and 2.398 (2dd, 4.4 and 12.8 Hz, H-3eq^{5,6}), 1.863 and 1.708 (2s, 2 x Ac), 1.104 (s, tBu)] from known compound **6**¹⁰ in 3 steps (i Ph₃SnH, AIBN in PhH, ii NaOMe, MeOH-PhMe, iii CH₂N₂ in 39% overall yield). All of the attempted glycosylations of **5** (R=Bn) as well as **6** with **3** (X=SMe, Br, F, and OCNHCCl₃) in our hands occurred at NHAc groups linked to either residue **5** or **6** but not at OH group of residue **2** of **5** and **6**, resulting in the formation of only acid labile imidates.



Scheme 3

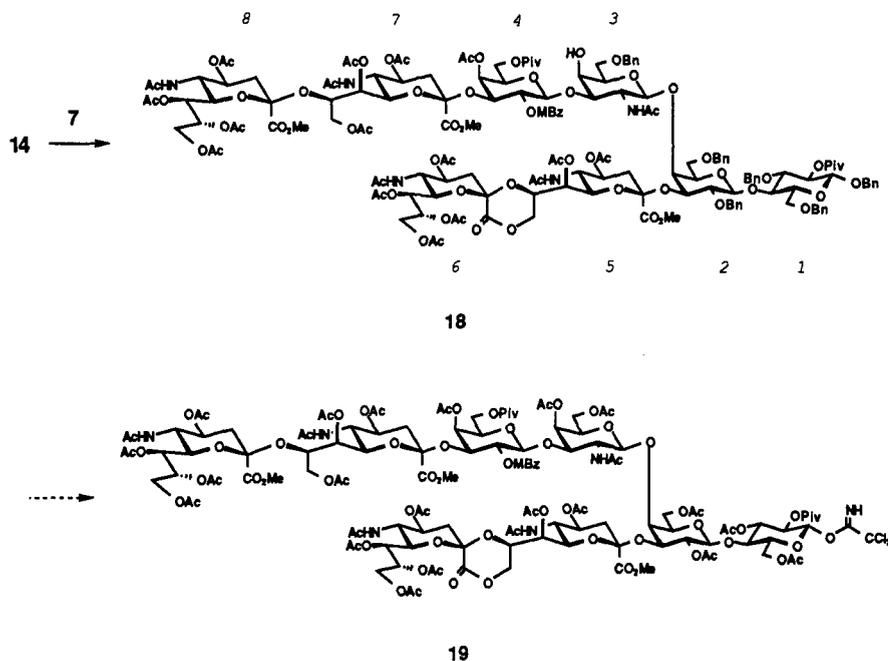
In order to lower the nucleophilicity of NHAc groups in compound **5** (R=Bn) toward glycosyl donors, we had to change the protective group of sialic acid residues from benzyl to electron withdrawing acetyl groups. According to this scenario, we studied compound **5** (R=Ac) as a glycosyl acceptor. To our delight, the glycosylation of **5** (R=Ac, ref 11) with **11** went smoothly¹² in the presence of PhSeOTf in CH₃CN to give an 89% of **12**: [α]_D +6.7° (c 0.6, CHCl₃); R_F 0.43 in 20:1 CHCl₃-MeOH; δ_H 6.235 (s, CHPh), 5.306 (d, 8.8 Hz, H-1³), 2.699 and 2.671 (2dd, 4.4 and 12.8 Hz, H-3eq^{5,6}), 1.744 (t, 12.8 Hz, H-3ax^{5,6}), 1.197 (s, tBu). Having successfully carried out a chain elongation at O-4 of residue **2** of

compound **5**, N-phthaloyl function of the protected pentasaccharide **12** was then converted into N-acetyl group in 4 steps (i LiI-Py, ii NH₂NH₂-H₂O, iii Ac₂O-MeOH, iv CH₂N₂, 95% overall) to give **13**: [α]_D -5.5° (c 0.9, CHCl₃); R_F 0.33 in 5:3 Me₂CO-hexane; δ_{H} 6.276 (s, CHPh), 5.117 (dd, 8.4 and 9.2 Hz, H-2¹), 3.996 (s, OMe), 2.242 (dd, 4.7 and 13.5 Hz, H-3eq^{5or6}), 1.789 (t, 12.8 Hz, H-3ax⁵), 1.169 (s, tBu). Treatment of **13** with 4:1 AcOH-H₂O afforded 87% of **14** which is ready for further chain elongation at O-3 of residue 3. Compound **14** had [α]_D -5.5° (c 0.3, CHCl₃); R_F 0.33 in 19:1 CHCl₃-MeOH; δ_{H} 5.200 (ddd, 5.5, 11.3 and 16.5 Hz, H-4⁶), 5.135 (dd, 8.1 and 9.2 Hz, H-2¹), 5.024 (ddd, 5.1, 12.1 and 16.8 Hz, H-4⁵), 2.346 (dd, 5.5 and 14.0 Hz, H-3eq⁶), 2.240 (t, 14.0 Hz, H-3ax⁶), 2.196 (dd, 5.1 and 14.3 Hz, H-3eq⁵), 1.716 (t, 14.3 Hz, H-3ax⁵), 1.170 (s, tBu).



Scheme 4

The building block **2** was prepared¹⁰ starting from **15** (ref.10). Glycosylation of **16** with 0.66 equivalent of **15** in the presence of Hg(CN)₂-HgBr₂-MS4A in CCl₄ and subsequent acetylation gave 33% of **17**: δ_{H} 5.311 (d, 3.4 Hz, H-4⁴), 3.421 and 2.963 (2d, 10.3 and 10.6 Hz, H-3^{7,8}). Further conversion of **17** into **2** was achieved in 4 steps (i Ph₃SnH, AIBN in PhH, ii H₂, 10% Pd-C in MeOH, iii Ac₂O-Py-DMAP, iv MeSSnBu₃-SnCl₄, 17% overall). Compound **2** had R_F 0.38 in 20:1 CHCl₃-MeOH; [α]_D +29.3° (c 0.4



Scheme 5

in CHCl₃); δ_{H} 4.864 (d, 9.6 Hz, H-1⁴), 3.828 and 3.796 (2s, 2 x OMe), 2.365 (s, C₆H₅Me), 2.221, 2.183, 2.166, 2.165, 2.137, 2.047, 2.046, 2.045, 1.956, 1.915 and 1.829 (11s, 10 x Ac and SMe), 1.202 (s, tBu). Crucial coupling of **14** with 1 equivalent of **2** in the presence of PhSeOTf in (CH₂Cl)₂ afforded 16% of the designed octasaccharide **18** along with 67% of recovered pentasaccharide **14**. It is to be noted that reaction of trichloroacetimidate corresponding to **2** with **14** in the presence of either TMSOTf or BF₃•OEt₂ in the same solvent failed completely. Stereochemistry of the glycosylation was confirmed by the following ¹H NMR of the compound **18**: R_F 0.28 in 1:2 CHCl₃-THF; δ_{H} 5.395 (dd, 7.7 and 9.5 Hz, H-2⁴), 4.801 (d, 7.7 Hz, H-1⁴), 3.800, 3.785, and 3.785 (3s, 3 x OMe), 2.368 (s, PhMe), 1.177 and 1.162 (2s, 2 x tBu). The regiochemistry of the coupling was confirmed by ¹H NMR of the acetate of **18** which contained a deshielded signal for H-4³ at δ 5.420 (d, 3.7 Hz). The Compound **18** could be regarded as a plausible precursor for the imidate **19**.

In summary, a first synthesis of properly protected octasaccharide **18** which was regarded a key intermediate for the synthesis of GQ1b was achieved by controlling the reactivity of OH-4 of residue 2 relative to NHAc of residues 5 and 6 in compound **5** by changing the protective groups of hydroxyl functions from benzyl to acetyl. Crucial coupling between pentasaccharide block **14** and trisaccharide block **2** was finally executed in the presence of thiophilic promoter.

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